

# Effects of different temperatures and water stress in germination and initial growth of creole genotypes of maize from three different agroclimatic regions of San Luis Potosí (México)

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**Keywords:** abiotic stress, drought, heat, climate change, polyethyleneglycol

## Abstract

Most of the studies on the effects of climate change on maize have been carried out on grain yield; few have been centred on germination and growth. In this investigation, experiments were carried out at different temperatures and water stress to evaluate the germination and initial growth of 12 creole maize genotypes from different agroclimatic regions of San Luis Potosí. The temperatures were 20, 25, 30 and 35 °C, proposed for having relation with the scenarios of temperature increase due to climate change in San Luis Potosí. The simulation of the water stress was carried out by the use of PEG-8000 (-0.5, -1 and -1.5 MPa), a control (0 MPa) was used. Four repetitions of 25 seeds for each creole genotype were carried out under a design at random with factorial arrangement. Germination and initial growth of the genotypes were negatively affected by the increase of temperature and decrease of water stress. Also these results were depended by the genetics materials origins and the climatic characteristics of each agroclimatic region of the state. In addition, the genotypes from less conditions of stress (Huasteca) were the most affected in comparison to materials from unfavorable conditions for the growth of maize plants (regions Altiplano and Media). Results suggest that the particular conditions of each region has influenced in the dynamic of germination and growth. We concluded, the related effects of climate change have affected differently and negatively the germination and initial growth of early maize from different agroclimatic regions.

## Introduction

In Mexico, climate change can cause an increment of the average annual temperature from 1 to 4 °C and changes in the average yearly precipitation from 0.4 to 1.4 times during the present century, in function of the scenario of the population growth (IPCC, 2007). Climate change is a modification in the state of the climate that can be identified (e.g. using statistical tests) by changes in the mean and/or the variability of its properties and that persists for an extended period, typically decades or longer. It refers to any change in climate over time, whether due to natural variability or as a result of human activity (IPCC, 2013).

Temperature and water supply are critical drivers for seed (initiation, break) and germination. Hence, global

climate change is altering these environmental cues and will preclude, delay, or enhance regeneration from seeds, as already documented in some cases. Along with compromised seedling emergence and vigour, shifts in germination phenology will influence population dynamics, and thus, species composition and diversity of communities (Walck et al., 2011a). A scenario of rising temperatures, declining rainfall, increase in extreme weather conditions, and shifting pest and disease patterns will lead to more short-term crop failures and long-term production declines (Kang and Banga 2013; Chauhan et al. 2014). The increase in temperature and the intensification of droughts due to the consequence of climate change have brought negative effects in ecophysiological processes such as germination and establishment of seedlings (Walck et al., 2011b; Ara-

gon-Gastelumet et al., 2017). Recent evidence shows that maize (*Zea mays* L.) is relatively more sensitive to high temperature and drought stress during sensitive stages of gametogenesis, flowering, and early grain filling stages of crop development (Prasad et al., 2020).

Maize is the most important cereal cultivated in Mexico (FAO, 2018; García-Lara and Serna-Saldivar, 2019), and Mexico is considered the center of origin and domestication of maize and one of the most important centers in its diversity (Matsuoka et al., 2002). The genetic variability of the maize constitutes a wealth for the population and it can be the base to achieve the domestic alimentary sovereignty, especially in the face of the changes in climate (Preciado Ortiz and Montes Hernandez, 2011). According to Cabrera et al. (2002), 76.5% of the Mexican farmers use creole seeds for its cultivation and that use depends mainly on the type of agriculture; however, in the regions with typical rural agriculture 80 to 100% of the farmers use creole seed fundamentally for its production. However, their production is generally limited for a series of factors like drought, plagues and diseases among others that represent economic losses particularly for smallholder farmers (Bergvinson, 2004).

The state of San Luis Potosí (S.L.P.) has been considered as a potential source of genetic variation in maize. Fourteen variants of maize have been documented in a study (Ávila-Perches et al., 2010a). The high abundance of the maize variants in the state can be due to wide diversity of the surroundings, in way similar to what happens in the Mexican territory. This way, it is possible to recognize different agroecological regions, from the warm and humid climatic conditions to the dry and hot or temperate ones (INEGI, 2012). Nevertheless, it is probable that the abundance of the maize variants in the state are much bigger that consigned by Ávila-Perches et al. (2010b) because just in three towns of the Huasteca, a region that represent a fraction of the whole territory of S.L.P., 11 different variants were registered (Heindorf et al., 2019).

It is proposed that the temperature values in S.L.P. will increase for a short-term scenario (2010-2040) and long term (2070-2100) (Medellín-Milan et al., 2006; Ávalos Lozano et al., 2017a). Also, the variation in the quantity of rain received in a month through the successive years is very big. Algara Siller et al. (2009) and Espinoza Jiménez & Gómez Mendoza (2019a) have also indicated the presence of drought in the state of S.L.P. It is probable that such an increase in temperature promotes, or associate with a decrease in the quantity of rain, will impact in the availability of water in soil and it will generate modifications like changes in the agricultural calendar, phenological alterations, and decrease of

yields, like it has been documented in Sinaloa in crop production of maize, bean (*Phaseolus vulgaris*), potato (*Solanum tuberosum*), red tomato (*Solanum lycopersicum*) among others (Ojeda-Bustamante et al., 2011). In the state of S.L.P. nearly 14,000 ha of maize are cultivated annually, of which a high proportion, around 80%, is cultivated under dry conditions (SIAP, 2017). Hence, the production of maize depends in great measure of the tolerance from the adapted genotypes of maize to conditions variables and many extreme times from the climate. This fact can mean the existence of highly adaptable creole genotypes of maize to adverse environmental conditions, what means an opportunity of use of such phylogenetic resources.

The study of the results of creole genotypes of maize coming from different conditions can help to determine and understand how the species of maize adapt to environmental conditions they face during their development and establishment, which is necessary to predict some of the effects related to climate change in the abundance and distribution of species (Dávila et al., 2013). Also, the abundance of the breed and/or creole genotypes of maize and their adaptations to diverse environmental conditions could represent alternatives to cultivate in scenarios that are poised to occur due to effects of climate change. The objective of this work was to evaluate the germination and initial growth of creole genotypes of maize coming from different agroclimatic regions at different temperatures and water stress (drought) under laboratory conditions. The above-mentioned, with the hypothesis that germination and initial growth of creole genotypes adapted to specific local conditions respond differently and in dependence of their origins when exposed to related effects of climate change.

## Material and methods

### Selection of creole genotypes of maize

The materials evaluated were collected in the state of S.L.P., in which, with basis on the mean annual of temperature and precipitation; three areas were identified whose average temperature and precipitation are around 14.5, 18.5 and 22.5 °C; 400, 700 and 1200 mm respectively (Noyola-Medrano et al., 2009a). Such areas were denominated *Altiplano*, *Media* and *Huasteca* respectively, and some features are described (Figure 1 and Table 1). A total of 37 distributed samples were obtained in the following way: 10 from the region of *Altiplano*, 11 from *Media* and 16 from *Huasteca*.

The collected samples were assessed as established by Carballo & Benítez (2003). This way a total of 14 variables were described: longitude (cm), diameter (mm) and conicity of cob, number of rows per cob, number

**Table 1 - Denomination of evaluated genotypes, climatic characteristics and futures scenarios of temperature based on the mean annual temperatures (M.A.T.) of the three regions from the state of San Luis Potosí, México.**

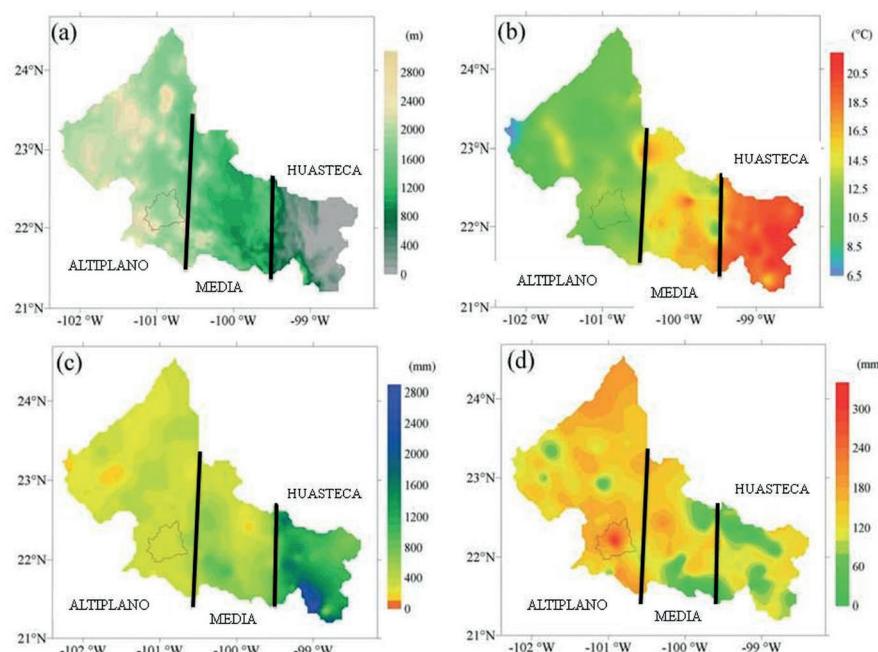
Genotypes	Region	Predominate climate based on modifications of the Köppen climate classification system and M.A.T.	Future mean annual temperature (M.A.T.) in short term (2010-2040) (M.A.T. + 0.75-1 °C)	Future mean annual temperature (M.A.T.) in long term (2070-2100 M.A.T. + 2.75-3.0 °C)
A3				
A4				
A8	Altiplano	BS1kw(e)gw" BSohw(e) gw"14.5 °C	15.25-15.5 °C	17.25-17.5 °C
A9				
A10				
M2				
M4	Media	Cb(w2)(w)(l') 18.5 °C	19.25-19.5 °C	21.25-21.5 °C
M11				
H4				
H9	Huasteca	(A)Cam(f)(e)gw" Am(e)gw" 22.5 °C	23.25-23.5 °C	25.25-25.5 °C
H10				
H11				

References: García, 2004. IPCC, 2013b; Ávalos Lozano et al., 2017b.

of grains per row, row arrangement, color and grain type, dry weight of 100 seeds (g), volume (mL) of 100 seeds in 50 mL, longitude (mm); thickness (mm) and wide (mm) of grain, phenological term (months). On the previous data were added, data on thermal and rainfall conditions facilitated by the producer. Such information was systematized with the following scale:

low, regular and high.

With the objective of containing the samples in conglomerates (clusters) with high grade of internal homogeneity and external heterogeneity among the samples (Sanjinez and Julca, 2019), and to select representative samples from the all collected, a multivariate analysis was carried out to form groups. For



**Fig. 1 - Horizontal distribution of (a) field elevation, (b) annual mean temperature, (c) annual mean precipitation, (d) annual mean evaporation of the three agroclimatic regions of the state of San Luis Potosí. Adapted from Noyola-Medrano et al. (2009b).**

processing the statistical software Statistical Analysis System (SAS) 9 was used. The resulting groups of the cluster analysis used in this investigation were: white maize's of 5 months of life cycle (genotypes A3, A4 and A8), white maize of 3 months (genotype A9) and purple maize (genotype A10) of 4 months of the region of *Altiplano*; white maize of 4 months (genotype M2), purple maize of 3 months (genotype M4) and black maize of 3 months (genotype M11) of the region of *Media*, white maize's of 4 and 3 months (genotypes H4 and H11 respectively) and yellow maize's of 3 months (genotypes H9 and H10) of the region of *Huasteca*.

#### **Treatments of temperature and simulation of water stress**

Germination experiments were conducted in controlled conditions of constant temperature (20, 25, 30 and 35 °C) and water potential [0 (control), -0.5, -1.0, and -1.5 MPa]. The temperatures were proposed for having relationship with scenarios of increase temperature due to effects of climate change (Figure 1 and Table 1). The conditions of soil humidity were determined at different levels of water potential since a very big variation in the quantity of rain received in a month through the successive years is expected (Espinoza-Jimenez and Gómez-Mendoza, 2019b).

The simulation of water deficit in the substrates was carried out with polyethyleneglycol 8000 (PEG-8000) as solution, with the purpose of simulating variations in the availability of soil humidity (Villalobos and Peláez, 2001; Huang *et al.*, 2018). This solution was used because it has been documented that it doesn't present toxicity on seeds (Khalil *et al.*, 2001). A control was also used with water potential of 0 MPa from deionized water.

To know the quantity of necessary PEG-8000 to obtain the potential water desired, the formula proposed by Burlyn (1983) was used:

$$\text{PEG} = \frac{4 - (5.16 \varphi T - 560 \varphi + 16)^{0.5}}{(2.58T - 280)}$$

Where:

PEG: Kg of PEG per liter of deionized water

$\varphi$ : Required water potential in bars (1 bar = 0.1 MPa)

T: Temperature of prepared solution

The three water potentials (-0.5, -1 and -1.5 MPa) were obtained by mixing deionized water and 145, 290 and 400 g L<sup>-1</sup> of PEG-8000 respectively. Water potential of each solution was measured in psychometric chambers

(Wescor, Logan, UT, USA). Samples of the solutions were taken approximately 2 mL and were placed in plastic trays in circular way 40 mm of diameter by 12 mm deep. The measurements obtained allowed to verify the levels of the wanted potentials.

#### **Experimental Design**

The experiment included a total of 192 treatments that resulted to a factorial arrangement of 4 x 4 x 12, with four repetitions. The temperature factor with four levels (20, 25, 30 and 35 °C), water stress with four levels (0, -0.5, -1 and -1.5 MPa) and the last factor represented by the 12 creole genotypes. Each experimental unit consisted of 25 seeds for creole genotype and the experimental units were distributed at random in a design of complete blocks.

Before sowing, the seeds were disinfected submerging them during 3 min in a solution of commercial chlorine (hypochlorite of sodium to 10%) (Viloria and Méndez Natera, 2011) and were then washed with abundant water to eliminate the excess of chlorine. Later on, the seeds were placed on filter paper, previously humidified with the solution of corresponding PEG, inside a plastic tray of 14 cm of long, 18 cm of wide and 8 cm of height. At once, the seeds were placed under predefined temperatures inside a chamber for plant growth at a neuter photoperiod of 12 hours light with the different levels of water potential.

#### **Germination variables**

##### **Percentage of germination (PG)**

A germinated seed is considered as one which has a root length of more than or equal to 2 mm. PG was calculated using the following formula (Scott *et al.*, 1983):

$$PG = \frac{(\text{Number of germinated seeds})}{(\text{Number of seeds sowed})} \times 100$$

It was determined at 15 days after sowing, instead of the seven days indicated by the International Seed Testing Association (ISTA) methodology. Other investigators have evaluated the germination of maize at 12 days (Laynez-Garsaballet *et al.*, 2007a), at 13, 14 and 15 days (Zagal-Tranquillo *et al.*, 2015), and at 16 days (Badr *et al.*, 2020).

##### **Mean germination time (MGT)**

It was determined according to the formula (Labouriau, 1983) at 15 days after sowing:

$$MGT = (\sum n_i t_i) / \sum n_i$$

Where:

$n_i$  is the number of seeds germinated at day  $i$  (not the accumulated number, but the number corresponding

**Table 2 - Mean squares (MS) from the analysis of variance of the percentage of germination (PG) and the mean germination time (MGT) of creole genotypes of maize from the state of San Luis Potosi at 15 days after sowing.**

Factors	PG			MGT	
	DF	MS	F-value	MS	F-value
Genotype (G)	11	9.1***	32.7	3.82***	23.29
Water stress (WS)	3	69***	906.8	83.75***	509.51
Temperature (T)	3	3.2***	42.05	5.6***	34.1
G x WS	33	3.4***	4.07	0.62***	3.79
G x T	33	8.5***	10.11	1.37***	8.34
WS x T	8	2.4***	11.77	1.38***	8.42
G x WS x T	74	7.5***	4.02	0.65***	3.97

DF: Degree of freedom; \*\*\* highly significant ( $P < 0.001$ ) according to Tukey test.

to the  $i$ th observation)

$t_i$  is the period from the start of the experiment to the  $i$ th observation (day)

#### Variables of growth

The variables of growth were determined for 15 seedlings per experimental unit at 16 days after sowing in two levels of water potential (0 MPa (control) and -0.5 MPa).

#### Rate of growth (RG) (mm.day $^{-1}$ )

Rate of growth (RG) was defined as the increment in longitude of the longest leaf and it was calculated in a period of 10 days. The following equation was used (Del Pozo et al., 1987):

$$RG = (L2 - L1) / (T2 - T1)$$

Where:

$L1$  and  $L2$  are the longitudes of the seedlings at 5 and 15 days respectively while  $T1$  and  $T2$  are the previously mentioned times.

#### Root length (RL) (cm)

Root length (RL) was determined with a rule graduated in cm starting from the root of major longitude.

#### Dry weight of shoots (DWS) and root (DWR) (g)

The samples were weighed, then dried off in stove at 70°C during 72 hours (Laynez-Garsaball et al., 2007b) and weighed in an analytic balance (Ohaus Compass™).

#### Statistical analysis

The data were submitted to an analysis of variance, using the procedure PROC GLM of the program Statistical Analysis System (SAS, 2003). The model consists of three fixed factors, 'genotypes', 'temperatures' and 'water stress' and their interaction 'genotype x tem-

perature x water stress'. Significant differences among the treatment means were then analyzed using the Tukey tests. The effects were considered significant if  $P < 0.05$ . Prior to the analysis, data were checked for normality and log-transformation was used to correct them. The level of significance was set at 5%.

#### Results and discussion

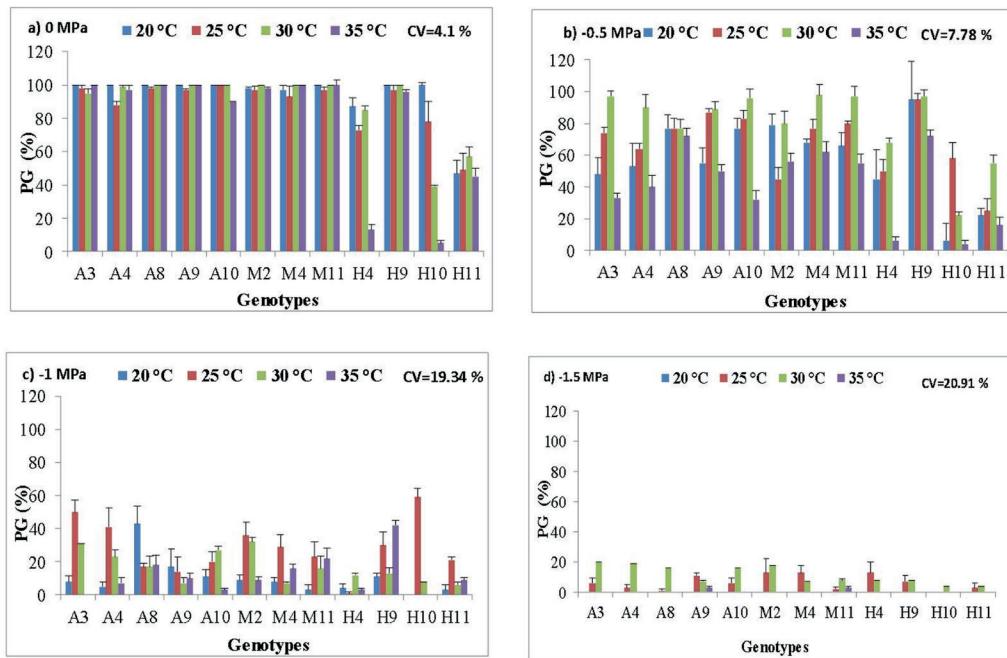
##### Germination variables

Based on the analysis of variance it was possible to observe significant effects of the genotypes, the water stress and temperatures. Also the interactions of the factors evaluated were significant ( $P < 0.001$ ) for the germination variables (Table 2).

##### Percentage of germination (PG)

The percentages of germination (PG) tend to reduce as water potential decreased; however, diminishing of PG varied depending on the creole genotypes and the temperatures (Table 2). At 0 MPa, the PG of genotypes from *Huasteca* was more affected by temperature variation than those from *Altiplano* and *Media*. The best germination percentages, with values highest than 96%, were registered in principally in genotypes from *Altiplano* and *Media*, even if genotypes A4 and A10 registered a reduced PG at 25 °C and 35 °C. The PG of the genotypes from *Huasteca* resulted highly reduced by variation of temperature in H4, H10 and H11 but in different way. In H4, 20-30 °C temperatures averaged a PG of 87% but it dropped to 13% at 35 °C, in H10 a gradual reduction of PG from 100% to 5% was registered along temperature increased. The PG of H11 was less than the half of all other genotypes (Figure 2a).

The temperatures promoted differentials reactions of PG in the water potential of -0.5 MPa. The genotypes



**Fig. 2 - Effect of different water potentials [a] 0,b) -0.5,c) -1,d) -1.5 MPa] and temperatures (20, 25, 30 and 35 °C) in the percentage of germination (PG) of creole genotypes of maize from different regions of the state of San Luis Potosi. A: Altiplano, M: Media, H: Huasteca. The vertical bars indicate the standard error (n=4).**

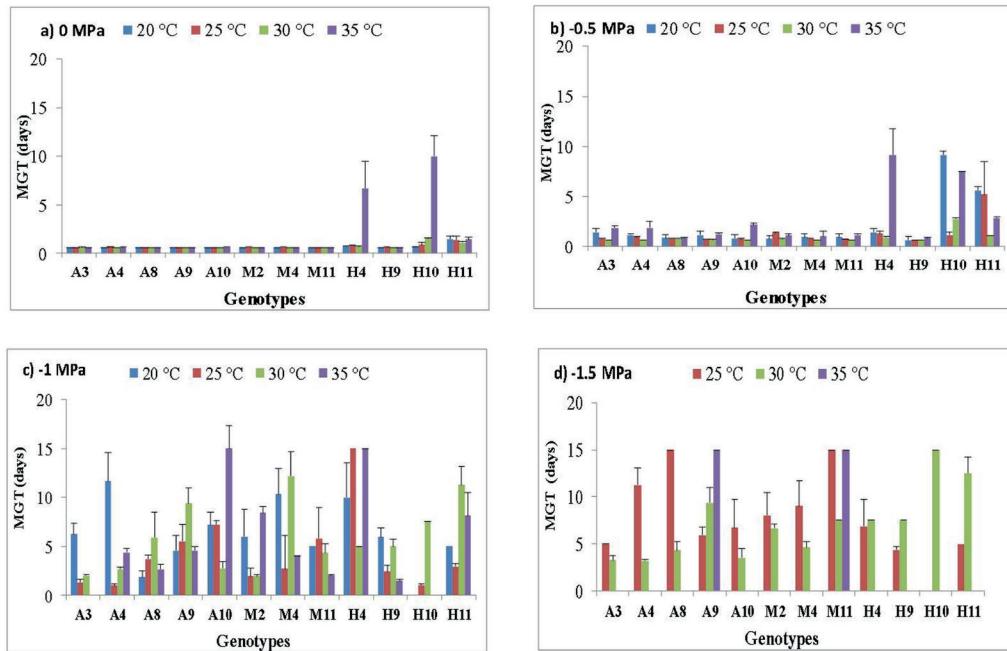
from *Altiplano*, *Media* and *H9* presented the highest PG at 25 and 30 °C. At 35 °C, the same genotypes reduced up to more than 50% their PG. The PG of the genotypes from *Huasteca* resulted highly influenced by variation of temperature in *H4*, *H10* and *H11*. *H4* and *H11* registered an increment of PG between 20 and 30 °C but drastically reduced at 35 °C until 6% and 16% respectively. However *H10* registered an average of 10.6% at 20, 25 and 35 °C while at 25 °C had the best PG with 58% (Figure 2b).

At -1 MPa, the PG of the creole genotypes of the *Media* was highly affected by the temperature factor while for those from *Altiplano* and *Huasteca* responded differently for temperature variation. In the same way, the creole genotypes from the *Media* registered the best PG at 25 °C with an average of 29% however those values decreased at the maximum temperature (35 °C) until 18%. For those from the *Altiplano* *A3* and *A4* registered the best values at 25 °C with an average of 46% while for the creole *A8*, *A9* and *A10* the best PG varied for each temperature. In addition the creole genotypes of the *Huasteca* were the most affected, specifically *H4* and *H11*. However *H9* and *H10* responded differently by the variation of temperature. For that *H9* registered best PG at 25 and 35 °C with 30 and 42% respectively (Figure 2c).

In a more extreme water stress (-1.5 MPa), germination was registered in all materials only at 30 °C although it was not more than 20%. At 25 °C, all of the materials

germinated although all below 15%. At 20 °C none of the genotypes germinated. At 35 °C, only the creole genotypes *A9* and *M11* registered a PG value of 3%, all the other materials did not register any germination (Figure 2d).

The germination response of creole genotypes of maize seeds was significantly affected by drought stress induced by PEG solutions and temperature. These findings extend those of Queiroz *et al.* (2019a) and Pawar *et al.* (2020), confirming that a water potential in a negative gradient tends to reduce the PG of maize seeds. In addition, the results noted in our study were related to temperature and creole genotypes. This study therefore indicates that creole genotypes from different regions responded differently for temperature; results were more accentuated at -1 and -1.5 MPa. Ramírez-Tobias *et al.* (2014) found that the germination of *Agave* species seeds decreased on dependence of species by up to 50% when water potential drops to -1.5 MPa. For the extreme stress (-1.5 MPa) at 20 °C, the maize creole genotypes did not germinate. Fancelli and Dourado-Neto (2000) stated that the temperature suitable for the germination of maize varies between 25 and 30 °C. Sans & Santana (2005) reported germination of maize may not occur at temperatures higher than 40 °C or near to 10 °C. Also, Borba *et al.* (1995) and dos Santos *et al.* (2019) reported that temperatures above 35 °C cause a significant decline in germination percentage of maize.



**Fig. 3 - Effect of different water potentials [a] 0, b) -0.5, c) -1, d) -1.5 MPa] and temperatures (20, 25, 30 and 35 °C) in the mean germination time (MGT) of the creole genotypes of maize from different regions of the state of San Luis Potosí. A: Altiplano; M: Media, H: Huasteca. MGT at 20 °C in d) did not exist because the creole genotypes did not germinate at this temperature and water potential (-1.5 MPa). Vertical bars show the standard error (n=4).**

#### Mean germination time (MGT)

The number of days for seeds to germinate increased when the water potential decreased; however, increment of the number of days for seed to germinate varied depending on the genotypes and temperatures. The genotypes from *Huasteca*, specifically H4 and H9, took more time to germinate at 35 °C and 0 MPa. In addition, all the others creole genotypes from *Altiplano* and *Media* took between 0.6 to 1.5 days to start germinating independently for the temperature (Figure 3a).

At water potential -0.5 MPa, the MGT varied by the creole genotypes and temperatures as in 0 MPa. In the same way, the most affected creole genotypes were from *Huasteca* but at 20, 25 and 35 °C. The maximum time to start germination was between 9.2-5.2 days. However the genotypes from *Altiplano* and *Media* responded equals no matter the temperature. Besides, they took between 0.6-2.2 days to start germinating (Figure 3b).

At -1 MPa, all the creole genotypes took more time to germinate than those observed at 0 MPa. These values were associated with creole genotypes and temperatures. For genotypes from *Huasteca*, the majority of them took less time to germinate at 25 and 30 °C between 1.5-2.5 days. In addition for genotypes from *Media* and *Altiplano*, most of them registered less time at 25, 30 and 35 °C however this response depended for each

genotype and took between 1.9-4.3 and 1.3-4.6 days respectively (Figure 3c).

At -1.5 MPa, the MGT varied depending creole genotypes and temperatures. Also they took more time to germinate in comparison to 0, -0.5 and -1 MPa. The genotypes did not germinate at 20 °C. In addition, all the genotypes from *Altiplano* and *Media* took less time to start germination at 30 °C with an average of 5-6 days while genotypes from *Huasteca* registered the less MGT at 25 °C with an average of 5 days (Figure 3d).

Our results reveal more negative is the water potential, longer it takes for the creole genotypes to germinate. This variable is important since it explains the time it takes for seeds to germinate after being embedded and thus take advantage of the short periods of rain, especially in arid and semi-arid areas (De la Barrera and Nobel, 2003). The results confirmed that the MGT dependent on the creole genotypes and temperatures (Tsougkrianis et al., 2009). According to Dürr et al. (2015) the base temperature for 241 species (including maize) to germinate were below 0 °C and below 20 °C and this results may due to the climatic conditions where the species grow or originated. Low temperatures delay germination, decrease reserve mobilization, and consequently the emergence velocity (Cruz et al., 2007). In the case of high temperatures, the biochemical processes of reserve mobilization are negatively af-

**Table 3 - Means squares (MS) from the analysis of variance of the growth variables of the creole genotypes of maize from the state of San Luis Potosí at 16 days after germination.**

Factors	DF	RG		RL		DWS		DWR	
		MS	F	MS	F	MS	F	MS	F
Genotype (G)	11	21.5*	49.2	0.26*	19.34	0.02*	29.16	0.05*	47.2
Water Stress (WS)	1	19.2*	43.9	5.64*	419.1	0.45*	451.4	0.41*	367.4
Temperature (T)	3	905.3*	2074.8	2.39*	177.8	0.24*	240.8	0.09*	84.8
G x WS	11	4.47*	10.25	0.15*	11.66	0.002*	1.85	0.02*	16.5
G x T	33	11.67*	26.76	0.18*	14.03	0.014*	14.38	0.03*	27.2
WS x T	3	92.45*	211.9	0.33*	24.73	0.009*	8.8	0.03*	29.3
G x WS x T	33	6.5***	14.96	0.07***	5.75	0.002*	1.81	0.006*	5.2

DF: degree of freedom, F: F-value; RG: rate of growth; RL: root length; DWS: dry weight of shoots; DWR: dry weight of root; \*, \*\*: significant at P<0.05 and P<0.001, respectively.

fected, due to low water availability. In addition, it may alter the constitution of cell membranes, affecting metabolic processes, especially cellular respiration (Taiz & Zeiger, 2004). For the factor water stress, Campos *et al.* (2020) recorded in succulents (*Agave lechuguilla* Torr. And *Agave salmiana* Otto ex Salm-Dyck) the time to reach maximum germination increases at more negative water potential. For corn germination in water stress conditions, the germination speed index expressed in seeds.day<sup>-1</sup> decreases as the water potential decreases (Khatami *et al.* 2015; Santos *et al.*, 2016).

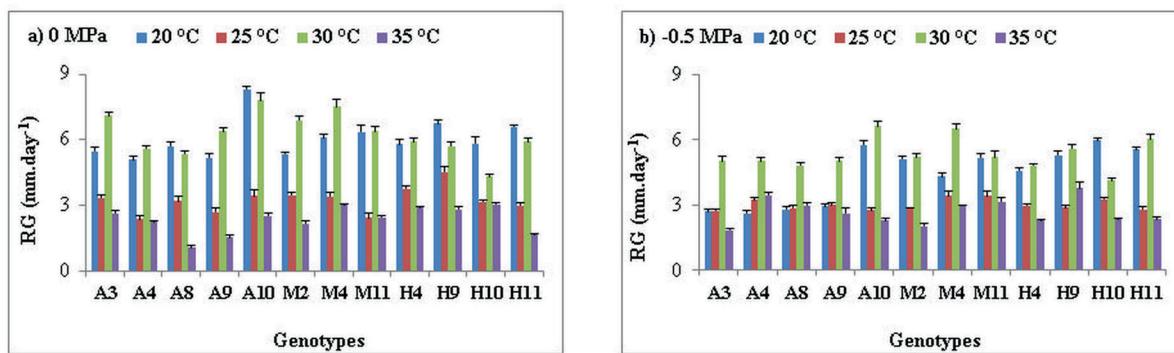
#### Variables of growth

The results of the analysis of variance showed significant effects (P < 0.05) for the main effects of creole genotypes, water potential levels and temperature, as well as significant interactions, for the variables evaluated. The significant interaction between the main effects of the factors (genotypes, water stress and temperature) indicates that genotypes have a distinct response when exposed to different drought levels and temperature (Table 3).

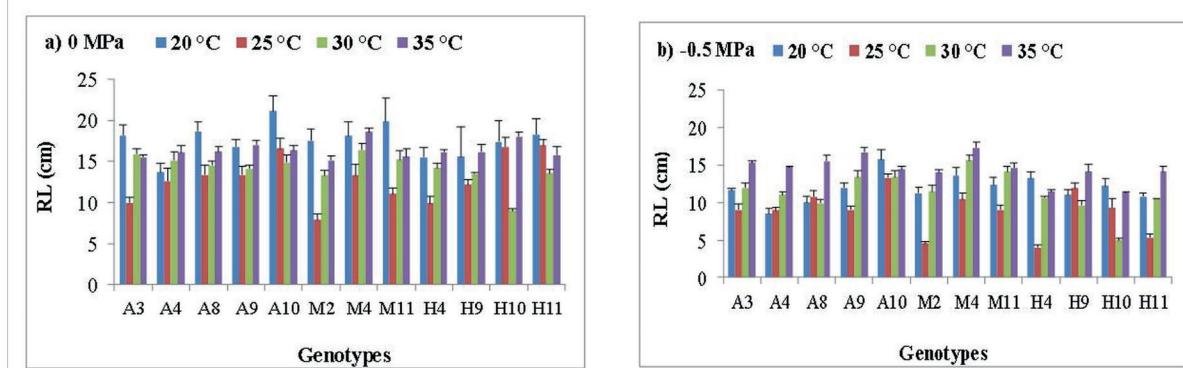
#### Rate of growth (RG)

High variation of the RG was registered among combination of temperatures, genotypes and water potential; however, tendencies due to reduction of water potential and temperature change were observed. Reduction of water potential promotes different patterns of RG values as temperature changed and a reduction of average RG. The RG of creole genotypes was drastically reduced with the decrease of water potential level from 0 to -0.5 MPa, in average of 30% respect to that obtained at 0 MPa. Maximum values of rate of growth (RG) were observed at 20 and 30 °C for all of the genotypes at 0 MPa and for genotypes from *Media* and *Huasteca* at -0.5 MPa. However, on genotypes from *Altiplano*, the maximum RG values only occurred at 30 °C. These maximum RG values doubled those registered at 25 and 35 °C (Figure 4). Then the seedlings growth velocity depended on the temperature and the origin of the genotypes.

According to Espinosa *et al.* (2015) the first process affected by the effect of water deficit is growth, which



**Fig. 4 - Effect of different water potentials [a] 0,b) -0.5 MPa] and temperatures (20, 25, 30 and 35 °C) on the rate growth (RG) of creole genotypes of maize from different regions of the state of San Luis Potosí. A: Altiplano; M: Media, H: Huasteca. Vertical bars indicate the standard error (n= 15).**



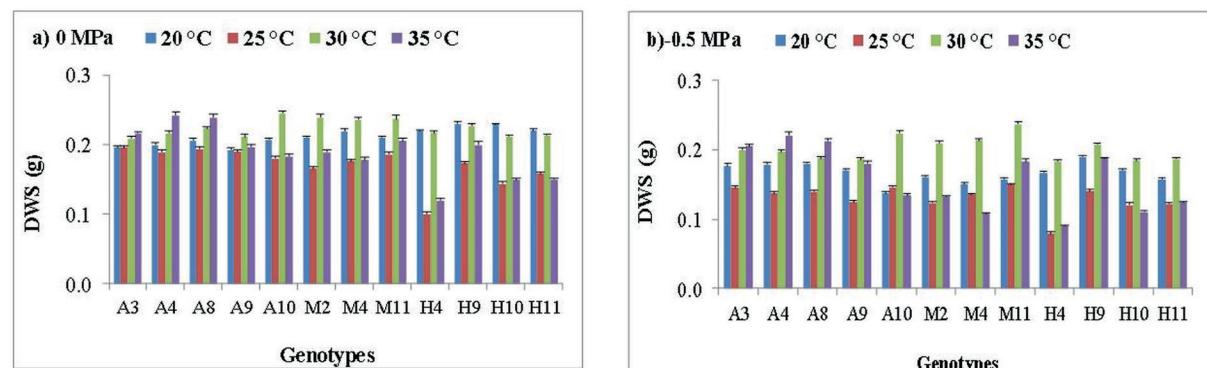
**Fig. 5 - Effect of different water potentials [a] 0,b) -0.5 MPa] and temperatures (20, 25, 30 and 35 °C) on the root length (RL) of creole genotypes of maize from different regions of the state of San Luis Potosí. A: Altiplano; M: Media, H: Huasteca. Vertical bars indicate the standard error (n= 15).**

begins with the germination, which comprises the imbibition, metabolism and growth start phases of the structures found in the embryo. Heat stress (27-35 °C during 14 days for maize (Virdi, 2016)) can induce irreversible damage to crop growth and yield (Fahad et al., 2017). It was found that a combination of drought and heat stress had a significant detriment effect on the growth in two contrasting maize landraces (Castro-Nava et al., 2014). Seedlings that emerge more quickly have higher time to develop compared to those that germinate later, and may have increased in seedling length (Queiroz et al., 2019b). Our results agree with this seedling growth dynamic because less growth was registered in germinated seedlings under water stress and high temperature (35 °C). Also at 25 °C the creole genotypes reduced their RG no matter the water potential and this effect can be associated to the temperatures that they are adapted during the initial growth contrary to the recommended temperature for corn growth as indicated Sun et al. (2017).

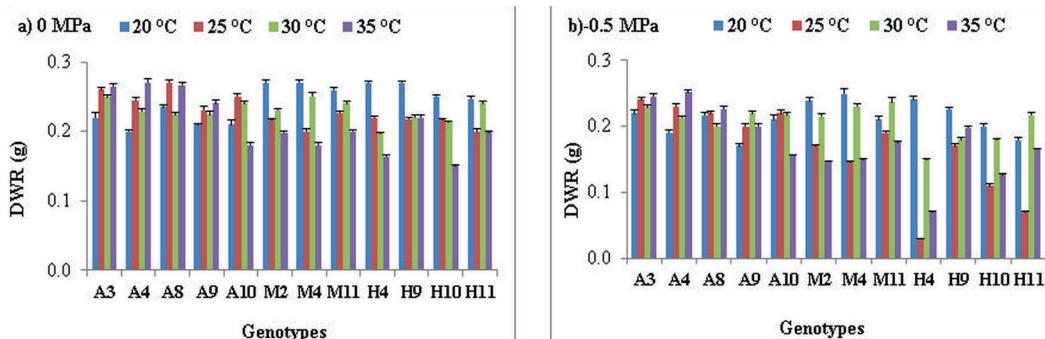
#### Root length (RL)

The root length of the maize genotypes was significantly affected by the combination of different temperatures and drought stress. That response presented differences depending on precedence of the creole genotypes. All the creole genotypes registered their maximum values of RL at 20 and 35 °C, but a fall of 21% at 25 and 30 °C in 0 MPa (Figure 5a). The minimum values of RL were observed in genotypes from *Altiplano* and *Media* in the two water potentials at 25 °C however for some genotypes from *Huasteca* the minimum RL moved at 30 °C. Reduction of water potential from 0 to -0.5 MPa decreased average RL to 35%. At -0.5 MPa, the RL at 20 °C tended to go down respect to 0 MPa (Figure 5b).

Trachsel et al. (2010) found that high temperature reduced RL of tropical maize seedlings. Similarly, Seiler et al. (1998) showed that increasing the root-zone temperature for sunflowers in growth pouches from 25 to 35 °C largely reduced length of the primary and lateral roots as well as the number of lateral roots. An increase in root-zone temperature from 22 to 38 °C for wheat grown hydroponically reduced the overall root length by 40



**Fig. 6 - Effect of different water potentials [a] 0,b) -0.5 MPa] and temperatures (20, 25, 30 and 35 °C) on dry weight of shoots (DWS) of creole genotypes of maize from different regions of the state of San Luis Potosí. A: Altiplano; M: Media, H: Huasteca. Vertical bars indicate the standard error (n= 15).**



**Fig. 7 - Effect of different water potentials [a) 0,b) -0.5 MPa] and temperatures (20, 25, 30 and 35 °C) on dry weight of roots (DWR) of creole genotypes of maize from different regions of the state of San Luis Potosí. A: Altiplano; M: Media, H: Huasteca. Vertical bars indicate the standard error (n= 15).**

to 60% depending on the level of tolerance of the evaluated genotypes (Tahir et al., 2008). Radhouane (2007) recorded a significant decrease in root length at -2 MPa with a reduction of more than 88% from Tunisian autochthonous pearl millet (*Pennisetum glaucum* (L.) R. Br.). Ren et al. (2020) showed that growth variables (root length, seedling, fresh weights of the root and aerial part decreased significantly under saline stress conditions in corn. Our results did not agree with this seedling growth dynamic where, in the combination of the abiotic factors, more growth of the RL was registered in germinated seedlings under water stress and at higher temperature (35 °C) in most of the genotypes. These results explain that genotypes with more roots at seedling stage subsequently developed stronger root system for the combination of the abiotic factors (high temperature and water stress), produced more biomass and had higher values for most characters determining seed yield (Bocev, 1963). However they decreased their RL in respect to control conditions.

#### Dry weight of shoots and roots

The DWS under control water potential (0 MPa) showed different responses of the genotypes for each temperature and agroclimatic region. In most of the genotypes from *Altiplano*, it was observed that the maximum values were found at 30 and 35 °C while 20 °C normally had low DWS (in average 80% respect to maximum values). A10 response was different, the temperatures 25 and 35 °C were not the ideal for his growth because it was significantly affected and their DWS decreased. Seedlings from *Media* and *Huasteca* registered their best DWS at 20 and 30 °C and their minimum ones at 25 and 35 °C; minimums were 22% and 32% reduced respect to the maximum values (Figure 6a).

The DWS of the genotypes was reduced with the decrease of water potential level but responses varied with genotypes. The genotypes from *Altiplano* (A3, A4, A8 and A9) registered their maximum values at 30 and 35 °C while A10 only at 30 °C. A3, A4, A8 and A9 reduced

drastically their DWS at 25 °C under water stress where they lost until 30% in comparison to 0 MPa. All the genotypes from *Media* obtained their maximum values at 30 °C, with 36% respect to those at 20, 25 and 35 °C. The genotypes from *Huasteca* registered their maximum values of DWS at 20 and 30 °C with 33% more weight for those found at 25 and 35 °C (Figure 6b).

As temperature increased, a slighted and discontinuous trend of DWR to increase was perceived in most of genotypes from *Altiplano* while in genotypes from *Media* and *Huasteca* the tendency was opposite and the water potential variation changed the trend (Figure 7). At 0MPa, most of the genotypes from *Altiplano* (A3, A4, A8 and A9) obtained their maximum values at 25 and 35 °C with 15% more to those registered at 20 and 30 °C. Also, genotypes from *Media* and *Huasteca* registered their best values at 20 °C but it was observed a decrease of 33% of their values with the increase of temperature (Figure 7a).

At -0.5 MPa, a reduction of the DWR occurred respect to 0 MPa. The genotypes from *Altiplano* delayed at least 10% of the weight at -0.5 MPa. Genotypes from *Media* were the most affected in the combination of 25 and 35 °C with -0.5 MPa. All the genotypes from *Media* significantly decreased their DWR and delayed at least 15% respect to 20 and 30 °C. Genotypes from *Huasteca* responded differently for each temperature at -0.5 MPa, they registered the maximum weight at 20°C but decreased up to 35% of their weight respect to 25, 30 and 35 °C. For *Huasteca* genotypes, the increase of the temperature reduced the DWR on H4 and H10 but increased on H9 and an absence of tendency was registered on H11 (Figure 7b).

In our investigation, the combination of each temperature (20, 25, 30 and 35 °C) with -0.5 MPa decreased the dry weights of the shoots and roots in most of the creole genotypes except in some from *Altiplano* (A3 and A10 at 20 °C, A9 and M11 at 30 °C). Ashagre et al. (2014) found different results in the evaluation of

highland maize cultivars under moisture stress at germination and seedling growth stages, they registered statistically higher values with the use of PEG-6000 (60, 120 and 180 mg. L<sup>-1</sup>) for the variables shoot and root dry weight. Farsiani & Ghobadi (2009), Khayatnezhad et al. (2010), Bibi et al. (2012), Radic et al. (2019), reported that drought drastically affected the fresh and dry weights of seedlings and roots in cultivars of sorghum, wheat, corn and sunflowers. These results agree with ours in the combination 20, 25 °C and water stress levels. However not in temperatures 30 and 35 °C; where creole genotypes A9 (at 30 °C), A8 and A9 (at 35 °C) were statistically equals. Toscano et al. (2017) recorded similar results in three sunflower cultivars at different water potentials (0, -0.15, -0.3, -0.45 and -0.65 MPa) at 20 °C. They found a significant decrease in root length, seedling length, fresh and dry seedling weights at the most negative water potential. Significant losses of germination and accumulation of dry matter were reported when using potentials of -0.5 MPa in creole of corn and beans (Aguilar-Benítez et al., 2014).

In most of the evaluated variables, the creole genotypes from *Huasteca* were the most affected and that results can be related to the particular conditions different to *Altiplano* and *Media* where they were collected. The region of *Huasteca* is the one with higher mean annual temperature and rainfall in comparison to *Altiplano* and *Media* (Figure 1); then their genetics materials are already adapted to no stress conditions. Then, these results can be associated to their local conditions where they grow with less stress in respect to those from the others regions of the state (Jiang et al., 1999). In addition, according to Mercer et al. (2008), maize landraces from tropical temperate conditions do not tolerate warmer climates due to local adaptation. In the state of San Luis Potosí, the sowing date of maize corresponds to the dates from May 15 to July 10 for *Altiplano*, May 15 to July 15 for *Media* while for the *Huasteca* it is from May 15 to 31 of July. On those dates, less monthly evapotranspiration, more cloudiness, more monthly average temperature and therefore more intensity in terms of the amount of water received in 24 hours in the *Huasteca* were recorded compared to the two others agroclimatic regions of the state. These environmental characteristics explain some of the responses obtained from the creole genotypes.

Similarly, maize is planted throughout Mexico, from very warm and humid climates at sea level to temperate and dry in the central plateau (equivalent to *Altiplano*). Distinct races and or genotypes of maize have been associated with particular environmental conditions since they were first classified (Wellhausen et al., 1952). Corral et al. (2008) classified Mexican maize ta-

king account rainfall, photoperiod and the most significantly temperature of their local adaptations or origins. These results have important implications for thinking about the effect of climate change adaptation of maize in the country because they indicated the way to take for contrasting the negative effects of climate change taking account the local conditions. Most notably, this is the first study to our knowledge to investigate effects of the increase of temperature and drought in creole genotypes of maize in San Luis Potosí taking account the three agroclimatic regions.

With the combination of the heat and water stress, the results obtained were null or below 20% for PG. The MGT was delayed by the lower osmotic potential and the different temperature while for the growth variables most of them decreased under water stress conditions and depended to temperatures. However at high temperature and water stress (35 °C/-0.5 MPa) the results decreased compared at 20, 25 and 30 °C.

Additionally, it has been documented that the effects of climate change will affect the grain yield of corn crops (Mina et al., 2012; Guerrero-Jiménez, 2016; Yang et al., 2017; Amouzou et al., 2019; Martins et al., 2019; Chen & Pang, 2020; Ureta et al., 2020); in this work, some of the effects related to climate change (drought and high temperatures) drastically affected the germination and initial growth of creole genotypes of maize from the state of S.L.P. under laboratory conditions. Then in this work, it has documented how the increase of temperature and the decrease of precipitation will affect negatively the germination and initial growth of creole genotypes from different agroclimatic regions. Besides the local particular conditions of each agroclimatic region has influenced the dynamic of germination and growth of the creole genotypes. Also, the materials were selected based on successful results from different environments, with different levels of water and thermal stress. Therefore, materials from less stressed areas (in this case *Huasteca*) will be less resilient to the related effect of climate change.

### Conclusions

The germination and initial growth of creole genotypes of maize coming from different agroclimatic regions were drastically affected by the increase of temperature and drought. The creole genotypes from the region with more high mean annual temperature and precipitation (*Huasteca*) were the most affected and this is associated to the local conditions adaptations of their genetic material with less conditions of stress in comparison to the materials from *Altiplano* and *Media*, regions of unfavorable conditions are associated with stress for the combination of less mean annual

temperature and precipitation. The creole genotypes from *Altiplano* and *Media* can represent an important phylogenetic potential to counteract negative effects of the increase of temperature and drought in the state of S.L.P for being the less affected in laboratory conditions and coming from unfavorable conditions for the development of maize plants.

### Acknowledgements

This work was supported by a Doctoral Research Fellowship award (2018-000012-01NACF-03824) from National Council for Science and Technology of Mexico (CONACYT) to the first author. To Drs. Cesar Iizaliturri Hernández, Guillermo Espinoza Reyes and the members of his laboratory for their supports during the experiments. To MPA Emmanuel Martínez Castro (Operational Unit Manager of National Seed Inspection and Certification Service (SNICS)-San Luis Potosí) during seed collection and travels among the state. For all the farmers who proportionated seeds and information.

### References

Aguilar-Benítez G, Peña-Valdivia CB, Vega JR, Castro-Rivera R, & Ramírez-Tobías HM, 2014. Seed germination and early root growth in common bean and maize landraces and improved cultivars at different water stress levels. *International Journal of Applied Science and Technology*, 4 (4).

Algara Siller M, Contreras Servín C, Galindo Mendoza G, y Mejía Saavedra J, 2009. Implicaciones territoriales del fenómeno de la sequía en la Huasteca Potosina. *Espaciotiempo* 4 : 56-67.

Amouzou KA, Lamers JP, Naab JB, Borgemeister C, Vlek PL, & Becker M, 2019. Climate change impact on water-and nitrogen-use efficiencies and yields of maize and sorghum in the northern Benin dry savanna, West Africa. *Field Crops Research*, 235, 104-117. <https://doi.org/10.1016/j.fcr.2019.02.021>

Aragón-Gastélum JL, Badano E, Yáñez-Espinosa L, Ramírez-Tobías HM, Rodas-Ortiz JP, González-Salvatierra C, & Flores J, 2017. Seedling survival of three endemic and threatened Mexican cacti under induced climate change. *Plant Species Biology*, 32(1), 92-99.

Ashagre H, Zeleke M, Mulugeta M & Estifanos E, 2014. Evaluation of highland maize (*Zea mays* L.) cultivars for polyethylene glycol (PEG) induced moisture stress tolerance at germination and seedling growth stages. *Journal of Plant Breeding and Crop Science*, 6(7), 77-83.

Ávalos Lozano JA, Magaña Rueda VO, Domínguez Cortinas G, Hernández Cendejas G, Díaz Gómez O, Ávila Galarza A, RoldánOrtíz M, Cruz González M, Portilla Rivera E, LoredoOsti C, Izaguirre Hernández JJ, Bonomi Cervantes MO, 2017. Programa Estatal de Acción ante el cambio climático del estado de San Luis Potosí. UASLP/SEGAM/Agenda ambiental/Variclim. [https://slp.gob.mx/SEGAM/Documentos%20compartidos/ESTUDIOS%20PROGRAMAS%20Y%20PROYECTOS/PEACC%202017%20SLP%20PRELIMINAR\\_V11A.pdf](https://slp.gob.mx/SEGAM/Documentos%20compartidos/ESTUDIOS%20PROGRAMAS%20Y%20PROYECTOS/PEACC%202017%20SLP%20PRELIMINAR_V11A.pdf)

Ávila-Perches MA, Dorantes-González JRA, Gámez-Vázquez HG, Gámez-Vázquez AJ, 2010. Conocimiento de la diversidad y distribución actual del maíz nativo y sus parientes silvestres en México, segunda etapa 2008-2009. 25 aniversario. Ciencia y Tecnología para el campo mexicano.

Badr A, El-Shazly HH, Tarawneh RA, & Börner A, 2020. Screening for Drought Tolerance in Maize (*Zea mays* L.) Germplasm Using Germination and Seedling Traits under Simulated Drought Conditions. *Plants*, 9(5), 565.

Bergvinson DJ, 2004. Opportunities and challenges for IMP in developing countries. In: Koul O, Dhalwal GS, Cuperus GW, (Eds), *Integrate Pest Management Potential, Constraints and Challenges*. CAB International, Wallsgrove, UK, pp. 281-312.

Bibi A, Sadaqat HA, Tahir MHN, & Akram HM, 2012. Screening of sorghum (*Sorghum bicolor* var Moench) for drought tolerance at seedling stage in polyethylene glycol. *J. Anim. Plant Sci*, 22(3), 671-678.

Bocev BV, 1963. Maize selection at an initial phase of development. *Kukuruzu*, 1: 54.

Borba CS, Andrade RV, Azevedo JT, Andreoli C, & Purcino AAC, 1995. Germinação de Sementes de Diversos Genótipos de Milho Tropical (*Zea mays* L.) em Diferentes Temperaturas. *Revista Brasileira de Sementes*, 16(2), 141-144. <https://doi.org/10.17801/0101-3122/rbs.v17n2p141-144>

Boyday M, Dirik H, Tilki F, and Çalikolu M, 2003. Effects of Water Stress on Germination in Six Provenances of *Pinus brutia* Seeds from Different Bioclimatic Zones in Turkey. *Turk J Agric For*. 27(2003). 91-97.

Burlyn EM, 1983. Evaluation of the water potentials of solutions of polyethylene glycol 8000 both in absence and presence of other solutes. *Plant Physiol*. 72:66-70. <https://doi.org/10.1104/pp.72.1.66>

Cabrera BEH, Macías-López A, Ruiz RD, Ramírez MV, Alvarado AD, 2002. Uso de semilla criolla y caracteres de mazorca para la selección de semilla de maíz en México. *Rev. Fitotec. Mex.*, 25, 17-23.

Campos H, Trejo C, Pena-Valdivia CB, García-Nava R, Conde-Martínez FV, Cruz-Ortega MdR, 2020. Water Availability Effects on Germination, Membrane Stability and Initial Root Growth of *Agave lechuguilla* and *A. salmiana*, *Flora*: <https://doi.org/10.1016/j.flora.2020.151606>

Carballo CA, & Benítez VAA, 2003. Manual gráfico para la descripción varietal del maíz (*Zea mays* L.). SAGARPA, SNICS. Colegio de Postgraduados en Ciencias Agrícolas. Montecillo, México. 69 p.

Castillo AM, Márquez MHR, Nieto CRM, and Tristán SSJ, 2003. Germination of locoweed seed (*Astragalus mollissimus* Torr) at different temperatures ranges and water stress levels. *Téc Pecu Méx*; 41(1): 85-89.

Castro-Nava S, Ramos-Ortiz VH, Reyes-Mendez CA, & Huerta AJ, 2014. Grain yield, photosynthesis and water relations in two contrasting maize landraces as affected by high temperature alone or in combination with drought. *Maydica*, 59(2), 104-111.

Chauhan BS, Prabhjyot-Kaur G, Mahajan RK, Randhawa H, Singh and Kang MS, 2014. Global warming and its possible impact on agriculture in India. In *Advances in Agronomy*, vol. 123, edited by D. L. Sparks, 65-121. Burlington, VT: Academic Press.

Chen C, & Pang Y, 2020. Response of maize yield to climate change in Sichuan province, China. *Global Ecology and Conservation*, 22, e00893. <https://doi.org/10.1016/j.gecco.2019.e00893>

Corral J, Durán Puga N, Sánchez González J, Ron Parra J, González Eguiarte D, Holland J, Medina García G, 2008. Climatic adaptation and ecological descriptors of 42 Mexican maize races. *Crop Science* 48, 1502-1512.

Cruz HL, Ferrari CS, Meneghelli GE, Konflanz VZPD, Vinholes OS, & Castro MAS, 2007. Avaliação de Genótipos de Milho para Semeadura Precoce sob Influência de Baixa Temperatura. *Revista Brasileira de Sementes*, 20(1), 52-60. <https://doi.org/10.1590/S0101-31222007000100008>

Davila P, Tellez O, Lira R, 2013. Impact of climate change on the distribution of an endemic Mexican columnar cactus in the Tehuacan-Cuicatlán Valley, Mexico, *Plant Biosyst.* 147: 376-386.

De la Barrera E, & Nobel PS, 2003. Physiological ecology of seed germination for the columnar cactus *Stenocereus queretaroensis*. *Journal of Arid Environments*, 53: 297-306.

Del Pozo HA, García-Huidobro J, Novoa R, Villaseca S, 1987. Relationship of base temperature to development of spring wheat. *Exp. Agric.* 23: 21-30.

dos Santos HO, Vasconcellos RC, de Pauli B, Pires RM, Pereira EM, Tirelli GV, & Pinho EV, 2019. Effect of Soil Temperature in the Emergence of Maize Seeds. *Journal of Agricultural Science*, 11(1). URL: <https://doi.org/10.5539/jas.v11n1p479>

Dürr C, Dickie JB, Yang XY, & Pritchard HW, 2015. Ranges of critical temperature and water potential values for the germination of species worldwide: contribution to a seed trait database. *Agricultural and forest meteorology*, 200, 222-232. <http://dx.doi.org/10.1016/j.agrformet.2014.09.024>

Espinosa PN, Martínez SJ, Santos TSA, Cadena IP, 2015. Selección de variedades nativas de maíz (*Zea mays* L.) por su tolerancia a la germinación bajo presión osmótica. VI. Reunión Nacional de maíces nativos. *Acta Fitogenética*. Vol. 2 (1):4.

Espinosa Jiménez A, & Gómez Mendoza L, 2019. Incendios forestales y el fenómeno de sequía: el caso de San Luis Potosí, México. Tlalli. *Revista De Investigación En Geografía*, (1), 13-24. <https://doi.org/10.22201/ffyl.26832275e.2019.1.381>

Fahad S, Bajwa AA, Nazir U, Anjum AS, Farooq A, Zohaib A, Sadia S, Nasim W, Adkins S, Saud S, Ihsan MZ, Alharby H, Wu C, Wang D, and Huang J, 2017. Crop Production under Drought and Heat Stress: Plant Responses and Management Options. *Frontiers in Plant Science*. <https://doi.org/10.3389/fpls.2017.01147>

Fancelli AL, & Dourado Neto D, 2000. *Ecofisiología e fenología. Produção de milho* (pp. 21-54). Guaíba:Agropecuária.

FAO (Food and Agriculture Organization of the United Nations), 2018. FAOSTAT.: FAO. Rome, Italy. Resource Document. <http://www.fao.org/faostat>. (Accessed 03 April, 2020).

Farsiani A, & Ghobadi ME, 2009. Effects of PEG and NaCl stress on two cultivars of corn (*Zea mays* L.) at germination and early seedling stages. *World Acad. Sci. Eng. Tech.*, 57, 382-385.

García E, 2004. Modificaciones al sistema de clasificación climática de Köppen. Universidad Nacional Autónoma de México.

García-Lara S, & Serna-Saldivar SO, 2019. Corn

history and culture. In: Serna-Saldivar, S.O. (Ed.), Corn, Chemists, St. Paul, Minnesota, pp. 1-18 5.

Giménez L, 2012. Producción de maíz con estrés hídrico provocado en diferentes etapas de desarrollo. Agrociencia Uruguay. 16 (2). 92-102.

Guerrero-Jiménez ZD, 2016. Short-term drought responses by seedlings of three maizes from contrasting environments in Michoacán, Mexico. Maydica, 60(2), 1-8.

Heindorf C, Reyes-Agüero JA, Van'tHooft A, Fortanelli-Martínez J, 2019. Inter- and Intraspecific Edible Plant Diversity of the TénekMilpa Fields in Mexico. Economic Botany XX(X), 2019, pp. 1-16. <https://doi.org/10.1007/s12231-019-09475>

Huang, Y, Xiang J, & Yin K, 2018. Differential gene expression analysis of the *Coix* transcriptome under PEG stress. Maydica, 62(1), 9.

INEGI (Instituto Nacional de Estadística y Geografía), 2012. Regiones agropecuarias de San Luis Potosí: Censo Agropecuario 2007. ISBN 978-607-494-406-8.

IPCC (Intergovernmental Panel on Climate Change), 2007. Climate Change: the physical science basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge: Cambridge University Press. pp.1-22.

IPCC (Intergovernmental Panel on Climate Change), 2013: Glosario [Planton, S. (ed.)]. En: Cambio Climático 2013. Bases físicas. Contribución del Grupo de trabajo I al Quinto Informe de Evaluación del Grupo Intergubernamental de Expertos sobre el Cambio Climático [Stocker, T.F., D. Qin, G.-K. Plattner, M. Tignor, S.K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex y P.M. Midgley (eds.)]. Cambridge University Press, Cambridge, Reino Unido y Nueva York, NY, Estados Unidos de América.

Jiang C, Edmeades G O, Armstead I, Lafitte HR, Hayward MD, & Hoisington D, 1999. Genetic analysis of adaptation differences between highland and lowland tropical maize using molecular markers. Theoretical and Applied Genetics, 99(7-8), 1106-1119.

Kang MS, and Banga SS, 2013. Global agriculture and climate change. J. Crop Improv. 27(6): 667-692. <https://doi:10.1080/15427528.2013.845051>

Kataria S, Baghel L, & Guruprasad KN, 2017. Pre-treatment of seeds with static magnetic field improves germination and early growth characteristics under salt stress in maize and soybean. Biocatalysis and agricultural biotechnology, 10, 83-90.

Khalil SK, Mexal JG, Murray LW, 2001. Germination of soybean seed primed in aerated solution of polyethylene glycol 8000. Online J. Biol. Sci., 1: 105-107.

Khatami SR, Sedghi M, & Sharifi RS, 2015. Influence of priming on the physiological traits of corn seed germination under drought stress. Annales of West University of Timisoara. Series of Biology, 18(1), 1.

Khayatnezhad M, Gholamin R, Jamaatie-Somarin SH, & Zabihi-Mahmoodabad R, 2010. Effects of peg stress on corn cultivars (*Zea mays* L.) at germination stage. World Appl. Sci. J, 11(5), 504-506.

Kin AG, Mazzola MB, & Cenizo VJ, 2015. Seed germination and seedling growth of the geophytic *Pterocactus tuberosus* (Cactaceae). The Journal of the Torrey Botanical Society, 142(4), 283-291.

Labouriau LG, 1983. A germinação de sementes. Washington: Organização dos Estados Americanos. p. 174.

Layne-Garsaball JA, Méndez Natera JR, Mayz-Figueroa J, 2007. Crecimiento de plántulas a partir de tres tamaños de semilla de dos cultivares de maíz (*Zea mays* L.), sembrados en arena y regados con tres soluciones osmóticas de sacarosa. Idesia (Arica), 25(1), 21-36. <https://dx.doi.org/10.4067/S0718-34292007000100003>

Marinoni LDR, Zabala JM, Patiño J, & Pensiero JF, 2017. Efecto de la temperatura y salinidad en la germinación y crecimiento inicial de un material naturalizado de *Lotus tenuis* Waldst. & Kit. Revista FAVE - Ciencias Agrarias 16 (2).

Martins MA, Tomasella J, & Dias CG, 2019. Maize yield under a changing climate in the Brazilian Northeast: Impacts and adaptation. Agricultural water management, 216, 339-350. <https://doi.org/10.1016/j.agwat.2019.02.011>

Matsuoka Y, Vigouroux Y, Goodman MM, Sanchez J, Buckler E, Doebley J, 2002. A single domestication for maize shown by multilocus microsatellite genotyping. Proc. Nat. Acad. Sci. USA 99:6080-6084. <https://doi.org/10.1073/pnas.052125199>

Medellín-Milán P, Ávalos-Lozano JA, Magaña-Rueda VO, 2006. Programa de Acción Ante el Cambio Climático de San Luis Potosí. Agenda ambiental/SEGAM. San Luis Potosí. <https://>

www.gob.mx/cms/uploads/attachment/file/316914/PEACC\_SLPcompressed.pdf

Méndez NJ, PF Ybarra, & Merazo PJ, 2010. Germinación y desarrollo de plántulas de tres híbridos de maíz bajo soluciones osmóticas. V. Polietilenglicol. Rev. Tecnol. ESPOL-RTE 23(1):49-54.

Mercer K, Martínez-Vásquez Á, & Perales HR, 2008. Asymmetrical local adaptation of maize landraces along an altitudinal gradient. Evolutionary Applications, 1(3), 489-500.

Mina U, Bhatia A, & Kumar U, 2012. Response of maize and its pest *Chilo partellus* to ozone and carbon dioxide exposure. Maydica, 57(3), 183-187.

Muscolo A, Sidari M, Anastasi U, Santonoceto C, & Maggio A, 2014. Effect of PEG-induced drought stress on seed germination of four lentil genotypes. Journal of Plant Interactions, 9(1), 354-363. <https://DOI:10.1080/17429145.2013.835880>

Noyola-Medrano MC, Ramos-Leal JA, Domínguez-Mariani E, Pineda-Martínez LF, López-Loera H, & Carbajal N, 2009. Factores que dan origen al minado de acuíferos en ambientes áridos: caso Valle de San Luis Potosí. Revista mexicana de ciencias geológicas, 26(2), 395-410.

Ojeda-Bustamante W, Sifuentes-Ibarra E, Íñiguez-Covarrubias M, & Montero-Martínez MJ, 2011. Impacto del cambio climático en el desarrollo y requerimientos hídricos de los cultivos. Agrociencia, 45(1), 1-11.

Ojeda-Silvera CM, Murillo-Amador B, Reynaldo-Escobar IM, Troyo-Diéguez E, Ruiz-Espinoza FH, & Nieto-Garibay A, 2013. Estrés hídrico en la germinación y crecimiento de plántulas de genotipos de albahaca *Ocimum basilicum* L. Rev. Mex. Cienc. Agric. 4:229-241.

Pawar KR, Wagh SG, Sonune PP, Solanke SR, Solanke SB, Rathod SG, & Harke SN, 2020. Analysis of Water Stress in Different Varieties of Maize (*Zea mays* L.) at the Early Seedling Stage. Biotechnology Journal International, 24(1), 15-24. <https://doi.org/10.9734/bji/2020/v24i130094>

Prasad PVV, Djanaguiraman M, Stewart ZP, & Ciampitti IA, 2020. Agroclimatología de Maíz, Sorghum, and Pearl Millet. Agroclimatología: Linking Agriculture to Climate, 60, 201-241. <https://doi:10.2134/agronmonogr60.2016.0005>

Preciado Ortiz RE, Montes Hernandez S, 2011. Amplitud, mejoramiento, usos y riesgos de la diversidad genética de maíz en México. eds (No. 631.5233 PRE. CIMMYT).

Queiroz MS, Oliveira CE, Steiner F, Zuffo AM, Zoz T, Vendruscolo EP, & Menis FT, 2019. Drought stresses on seed germination and early growth of maize and sorghum. Journal of Agricultural Science, 11(2), 310-318. <https://doi.org/10.5539/jas.v11n2p310>

Radhouane L, 2007. Response of Tunisian autochthonous pearl millet (*Pennisetum glaucum* (L.) R. Br.) to drought stress induced by polyethylene glycol (PEG) 6000. African journal of biotechnology, 6(9).

Radić V, Balalić I, Jaćimović G, Nastasić A, Savić J, & Marjanović-Jeromela A, 2019. Impact of drought and salt stress on seed germination and seedling growth of maize hybrids. Genetika, 51(2), 743-756.

Ramírez-Tobías HM, Peña-Valdivia CB, Trejo CR, Aguirre JR, & Vaquera HH, 2014. Seed germination of *Agave* species as influenced by substrate water potential. Biological Research, 47, 1-9. <https://dx.doi.org/10.1186/0717-6287-47-11>

Rangel-Fajardo MA, Gómez-Montiel N, Tucuch-Haas JI, Basto-Barbudo DC, Villalobos-González A, & Burgos-Díaz JA, 2019. Polietilenglicol 8000 para identificar maíz tolerante al estrés hídrico durante la germinación. Agronomía Mesoamericana, 30(1), 255-266. <https://dx.doi.org/10.15517/am.v30i1.34198>

Ren Y, Wang W, He J, Zhang L, Wei Y, & Yang M, 2020. Nitric oxide alleviates salt stress in seed germination and early seedling growth of pakchoi (*Brassica chinensis* L.) by enhancing physiological and biochemical parameters. Ecotoxicology and environmental safety, 187, 109785. <https://doi.org/10.1016/j.ecoenv.2019.109785>

Rodríguez-Morales J, Guillén S, & Casas A, 2013. Consecuencias de la domesticación de *Stenocereus stellatus* en el tamaño de las semillas y en la germinación en un gradiente de estrés hídrico. Botanical Sciences, 91(4), 485-492.

Sanjinez F, Julca A, 2019. Caracterización de parcelas productoras de arroz (*Oryza sativa* L.) en Tumbes, Perú. Agroind. sci. 9(1): 67-75. DOI: <http://dx.doi.org/10.17268/agoind.sci.2019.01.09>

Sans LMA, & Santana DP, 2005. Cultivo do Milho-Clima e solo. Sete Lagoas: EMBRAPA Milho e Sorgo.

Santos MACD, Rego MMD, Queiroz MAD, Dantas BF, & Otoni WC, 2016. SYNCHRONIZING THE

in vitro GERMINATION OF *Psidium guineense* Sw. SEEDS BY MEANS OF OSMOTIC PRIMING. Revista Árvore, 40(4), 649-660. <http://dx.doi.org/10.1590/0100-67622016000400008>

SAS, 2003. Institute Inc. Statistical Analysis System user's guide. Version 9.1. Statistical Analysis System Institute, Cary (NC).

Scott WR, Appleyard M, Fellowes G, and Kirby EJM, 1983. Effect of genotype and position in the ear on carpel and grain growth and mature grain weight of spring barley. *J. Agric. Sci.* 100: 383-391.

Seiler GJ, 1998. Influence of temperature on primary and lateral root growth of sunflower seedlings. *Env. Exp. Bot.* 40:135-146.

SIAP (Servicio de Información Agroalimentaria y Pesquera), 2017. Sagarpa, México. [http://infosiap.siap.gob.mx:8080/agricola\\_siap\\_gobmx/ResumenProducto.do](http://infosiap.siap.gob.mx:8080/agricola_siap_gobmx/ResumenProducto.do). Consultado el 5/11/2017

Sun CX, Chen X, Cao MM, Li MQ, Zhang YL, 2017. Growth and metabolic responses of maize roots to straw biochar application at different rates. *Plant and soil*, 416(1-2), 487-502. DOI 10.1007/s11104-017-3229-6

Tahir ISA, Nakata N, Yamaguchi T, Nakano J, ALI AM, 2008. Influence of high shoot and root-zone temperatures on growth of three wheat genotypes during early vegetative stages. *J. Agr. Crop Sci.* 194: 141-151.

Taiz L, & Zeiger E, 2006. *Plant Physiology* (4th ed.). Sinauer Associates, Sunderland, MA.

Toscano S, Romano D, Tribulato A, & Patanè C, 2017. Effects of drought stress on seed germination of ornamental sunflowers. *Acta Physiologiae Plantarum*, 39(8), 184. <https://doi.org/10.1007/s11738-017-2484-8>

Trachsel S, Stamp P, & Hund A. 2010. Effect of high temperatures, drought and aluminum toxicity on root growth of tropical maize (*Zea mays* L.) seedlings. *Maydica*, 55(3), 249.

Tsoukrianis N, Peña-Valdivia CB, Trejo C, Molina JD, 2009. El potencial de agua del sustrato en la germinación de maíces con tolerancia y sensibilidad a la sequía. *Agricultura Técnica en México* 35(4):363-369.

Turk M, & Eser O, 2016. EFFECTS OF SALT STRESS ON GERMINATION OF SOME SILAGE MAIZE (*Zea mays* L.) CULTIVARS. *Scientific Papers-Series A, Agronomy*, 59, 466-469.

Ureta C, González EJ, Espinosa A, Trueba A, Piñeyro-Nelson A, & Álvarez-Buylla ER, 2020. Maize yield in Mexico under climate change. *Agricultural Systems*, 177, 102697. <https://doi.org/10.1016/j.agrosy.2019.102697>

Vaz-de-Melo A, Santos LDT, Finoto EL, dos Santos Dias DCF, & Alvarenga EM, 2012. Germinação e vigor de sementes de milho-pipoca submetidas ao estresse térmico e hídrico. *Bioscience Journal*, 28(5).

Vide JM, 2009. Conceptos previos y conceptos nuevos en el estudio del cambio climático reciente. *Investigaciones Geográficas (Esp)*, (49), 51-63. [fecha de Consulta 4 de Abril de 2020]. ISSN: 0213-4691. Disponible en: <https://www.redalyc.org/articulo.oa?id=176/17617034003>

Villalobos AE, Peláez DV, 2001. Influences of temperature and water stress on germination and establishment of *Prosopis caldenia* Burk. *Journal of Arid Environments*. 49:2, 321-328. <https://doi.org/10.1006/jare.2000.0782>

Viloria H, Natera JRM, 2011. Relationship among electrical conductivity, pH of soaked-water, seed germination and seedling growth of corn (*Zea mays* L.) under two experimental conditions. *Scientia Agropecuaria*, 2(4), 213-228.

Virdi SK, 2016. HEAT AND WATER STRESS IN PLANTS-A REVIEW. *Harvest (Online); Bi-Annual journal*, 2016(2), 15-42.

Walck JL, Hidayati SN, Dixon KW, Thompson K, & Poschlod P, 2011. Climate change and plant regeneration from seed. *Global Change Biology*, 17, 2145-2161. <https://doi.org/10.1111/j.1365-2486.2010.02368.x>

Wellhausen E, Roberts J, Roberts LM, Hernández E, 1952. *Races of Maize in México: Their origin, Characteristics, and Distribution*. Harvard University, Cambridge.

Yang C, Fraga H, Van Ieperen W, & Santos JA, 2017. Assessment of irrigated maize yield response to climate change scenarios in Portugal. *Agricultural Water Management*, 184, 178-190. <http://dx.doi.org/10.1016/j.agwat.2017.02.004>

Yuan Z, Wang C, Li S, Li X, and Tai F, 2014. Effects of different plant hormones or PEG seed soaking on maize resistance to drought stress. *Can. J. Plant Sci.* 94: 14911499.

Zagal-Tranquilino M, Martínez-González S, Salgado-Moreno S, Escalera-Valente F, Peña-Perra B, & Carillo-Díaz F, 2016. Producción de forraje verde hidropónico de maíz con riego de agua cada 24 horas. *AbanicoVet.*; 6(1):29-34.

Zalama MT, & Kishk AMS, 2017. Evaluation of Seed Germinability and Field Emergence Of Some Maize (*Zea mays*, L.) Hybrids Under Salinity Stress Conditions. *Journal of Plant Production*, 8(5), 649-656.